

Diagnostic accuracy of a near point-of-care HIV drug resistance test OLA-Simple: a field validation study in Kenya

N Panpradist 1,2*, IA Beck 3*, A Miller 1, AM Cash 1, J Campbell 1, SWA Stewart 1,3, PS Ruth 4, B Chohan 1,5, P Owiti 6, G Akinyi 7, N Nyakundi 7, VA Sewe 7, RS Madada 2, V Onwonga 2, S Akasa 2, K Yamashita 1, B Tran 1, EC Kline 1, J Vrana 1, G Thakur 1, JH Kotnik 1, J Sprecher 1, Q Wang 1, S Gilligan-Steinberg 1, J Henthorn 1, JK Liu 1, KL Tukei 1, M Samadpour 8, L Kingwara 2, N Bowen 2, V Opollo 7, LM Frenkel 1,3, L Abougi 9, E Klavins 1, P Oyaro 7, BR Lutz 1†, R Patel 1†

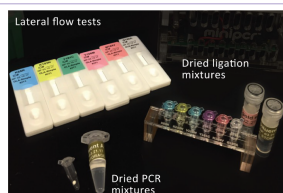
1 University of Washington, Washington, USA; 2 National HIV Reference Laboratory, Nairobi, Kenya; 3 Seattle Children's Research Institute, Washington, USA; 4 Stanford University, California, USA; 5 Kenya Medical Research Institute, Nairobi, Kenya; 6 University of Nairobi, Nairobi, Kenya; 7 Kenya Medical Research Institute / Center for Disease Control, Kisumu, Kenya; 8 IEH Consulting Group, Washington, USA; 9 University of Colorado, Colorado, USA * Authors with equal contributions † Co-corresponding authors (BRL: Engineering corresponding author, RP: Clinical corresponding author)



Presenting author
(nuttadap@uw.edu)

BACKGROUND

- Dolutegravir (DTG) is associated with improved antiretroviral therapy (ART) outcomes. Despite data suggesting that acquired HIV drug resistance (HIVDR) does not undermine efficacy of DTG-ART, **additional studies are needed to further evaluate the effects of extensive pre-existing nucleos(t)ide resistance on DTG-ART across subtypes.**
- In 2019, we reported development and clinical validation of "OLA-Simple," an easy-to-use, point-mutation HIVDR test. **1-3**



- Ready-to-go dried reagents - easy assay set up**
- Custom-designed lateral flow tests - visual readout**
- In-house software to guide "molecular biology novices"**

99.6% specificity
98% sensitivity
Validated in specimens from A, B, C, D, and AE subtypes

No training required
Guided by software, 1st-time users processed specimens with 97% accuracy.

~\$850 equipment
<\$20 reagents/ sample tested
to detect DR for NRTIs (TDF/3TC) and NNRTIs (EFV/NVP) from DNA/RNA

Same-day results
<4-hour total time
<10-min hands-on time

- This work presents:**
 - Our recent probe development to include **detection of HIVDR mutations against Abacavir**
 - Our **first-ever clinical validation of OLA-Simple** that is performed in laboratories from Kenya.

RESULTS AND DISCUSSION

PHASE I | On-site training local researchers using DNA controls



- Three and four technicians were trained at NHRL and KEMRI-CDC, respectively. During the observed training, US trainer observed the run performed by the local researchers. During the unobserved training, technicians operated independently without observation and relied completely on the software guide.
- Genotype classifications of the results obtained both during the observed and unobserved runs had a 100% agreements with the genotypes of the DNA controls.

PHASE II | Local researchers evaluated OLA-Simple on plasma specimens

- RT-PCR for OLA successfully amplified 134 specimens (91.8%) with plasma HIV RNA between 27 - 11013 copies/reaction (median: 317). The ones that failed had 35 - 1940 copies/reactions; thus RT-PCR failure could be due to a combination of factors:
 - Low RNA input (near the limit detection of RT-PCR)
 - PCR inhibitors carried from the extraction step, and/or
 - RNA degradation due to introduction of multiple freeze-thaw cycles of plasmas.
- From successfully amplified specimens, 132 had OLA-Simple image data for analysis of 924 codons which included 275 mutant, 612 wild-type and 37 (4%) indeterminate results (see Table 1).

Table 1. Summary of OLA-Simple results compared to Sanger with discordant results adjudicated by sensitive benchmark (laboratory OLA)

	K65R	L74V/I	K103N/S	Y115F	Y181C	M184V	G190A	Total	
True (-)	114	96	50	112	103	35	88	589	Sensitivity 94.7% [95%CI: 91.3-97.1]
True (+)	6	25	69	11	21	87	33	258	Specificity 96.3% [95%CI: 94.5-97.6%]
False (+)	2	3	8	0	2	2	6	23	
False (-)	0	0	4	4	1	4	1	14	
Indeterminate	10	8	1	5	5	4	4	37	Indeterminate 4% [95%CI: 2.8-5.5]

- OLA-Simple detected 6 low frequency mutant variants missed by Sanger (confirmed by laboratory OLA sensitive to 2% mutant) and misclassified 23 (2.4%) wild-type codons as mutant and 14 (1.6%) mutant codons as wild-type.
- Based on these results, OLA-Simple had **sensitivity of 94.7% and specificity of 96.3%.**

CONCLUSIONS AND NEXT STEPS

- This in-field validation study serves as a significant step towards implementation of OLA-Simple in LMICs. It reveals OLA-Simple's high sensitivity and specificity.
- Our next steps include validation of these probes on prospective samples, and developing OLA probes to detect DTG HIVDR mutations.

ACKNOWLEDGMENTS

We thank our collaborators: Dr. James Lai at the University of Washington, USA. Dr. Theresa Rossouw at University of Pretoria, SA; Dr. Gonzague Jourdain and Dr. Nicole Ngo-Giang-Huong at PHPT, Thailand; Dr. Jaime Soria at Hospital Nacional Dos de Mayo, Peru for stimulating discussion.

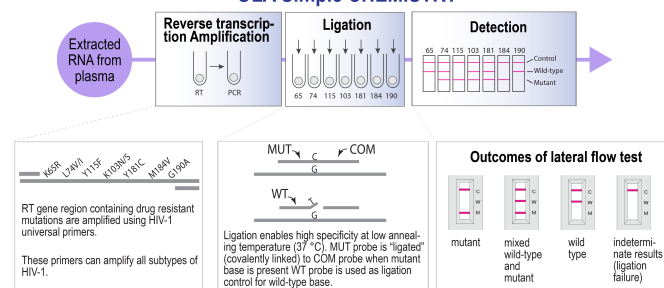


REFERENCES

- N Panpradist, et al. "OLA-Simple: A software-guided HIV-1 drug resistance test for low-resource laboratories." 2019. EBiomedicine.
- N Panpradist, IA Beck, et al. "Near point-of-care, point-mutation test to detect drug resistance in HIV-1: a validation study in a Mexican cohort." 2020. AIDS.
- J Vrana, N Panpradist, et al. "Implementation of an interactive mobile application to pilot a rapid assay to detect HIV drug resistance mutations in Kenya." 2022. PLOS Global Public Health.
- IA Beck, et al. "Rapid and sensitive oligonucleotide ligation assay for detection of mutations in human immunodeficiency virus type 1 associated with high-level resistance to protease inhibitors." 2002. Clinical Microbiology.

METHODS

OLA Simple CHEMISTRY



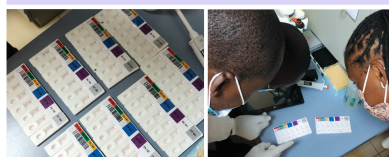
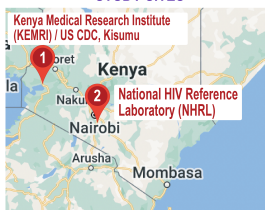
STUDY DESIGN

Study time frame: April-June 2022.

PHASE I (3 days on-site training)

Total of seven lab technicians performed OLA-Simple on two blinded control specimens (negative plasma spiked with DNA).

STUDY SITES



PHASE II 8-10 weeks of independent testing by local lab technicians

